Resistance of Bacillus Amyloliquefaciens Spores to Melt Extrusion Process Conditions

Abstract
With the increasing demand for functionalised textile materials, industry is focusing on research that will add novel properties to textiles. Bioactive compounds and their benefits have been and are still considered as a possible source of unique functionalities to be explored. However, incorporating bioactive compounds into textiles and their resistance to textile process parameters has not yet been studied. In this study, we developed a system to study the resistance of Bacillus amyloliquefaciens spores against melt extrusion process parameters, like temperature (21, 200, 250, 300 °C), pressure (0.1, 0.6 and 1.0 MPa) and residence time (0, 1 and 10 minutes). The spores were successfully embedded in PET (polyethylene terephthalate) films and fibres through melt extrusion. Afterwards the survival rate of the spores was determined after extrusion and the data was used to develop a quadratic equation that relates the survival rate to the spore concentration.

Key words: Bacillus amyloliquefaciens, spores, resistance, melt extrusion.

Introduction
Biotechnology is becoming increasingly important in the textile industry, leading to novel multifunctional textile products with unique values and functionalities in contrast to classical textile materials. Some of the new functional properties obtained in textiles and clothing materials via biotechnology include antimicrobial properties and enzymatic modification of textiles e.g. polyester, wool, cotton, etc. [1, 2]. Due to the growing need for multifunctional, eco-friendly and efficient textile products, the use of biologic-
spores during extrusion as a function of the temperature, pressure, residence time, shear stress and polymer used.

### Materials and method

#### Microorganism

Spores of the thermophile bacterial strain *Bacillus amyloliquefaciens* used were obtained from bio-mining in the Moroccan desert and a stock culture was cultivated at Ghent University (Biology Department). Spores of *B. amyloliquefaciens* are known to be resistant to high temperatures and pressures [13, 14], making them good candidates for extruding in polymers.

#### Resistance to temperature, pressure and residence time

To test the resistance of *B. amyloliquefaciens* spores to melt extrusion process parameters like the temperature, pressure and residence time, a special device was designed and developed at Ghent University (Biology Department). The spores were exposed to different test parameters based on a pilot study (Table 1).

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Values</th>
</tr>
</thead>
<tbody>
<tr>
<td>Temperature, °C</td>
<td>21 (Control), 200, 250 and 300</td>
</tr>
<tr>
<td>Pressure, MPa</td>
<td>0.1 (control), 0.6 and 1.0</td>
</tr>
<tr>
<td>Residence time, min.</td>
<td>0 (control), 1 and 10</td>
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</table>

Three different conditions were used: 1) temperature and atmospheric pressure, 2) temperature and pressure, 3) room temperature and pressure.

A compressor (HP3) was used to pressurise a known amount of dry spores in the sample holder, and then it was placed in a preheated dry air oven at the testing temperature required (Memmert UFE 500). A probe of thermocouple wire connected to a data logger (Testo 175-T3) was placed in direct contact with the sample in order to follow the actual temperature on the sample. When the temperature required was reached, the sample was left in that temperature for the specified amount of time, after which the sample was removed and left to cool down to room temperature [17].

Then the spores treated were inoculated by dissolving in 1000 µl of sterile physiological saline solution (0.8% NaCl) and diluted up to $10^{-15}$ fold. From this dilution, 50 µl was inoculated on a prepared culture medium (Nutrient Agar, Oxoid CM0003). The plates were incubated for 24 h at 40 °C. The survival of spores was determined by counting the colony forming units (CFU), after which a log calculation was made to obtain the results, expressed in log CFU/g dry weight.

The two-sample Kolmogorov-Smirnov statistical test was used to determine the significance of each test parameter on the number of resistant spores.

#### Extrusion of spores in PET films

Artnite thermoplastic PET pellets (Arntite A02307) from Royal DSM N.V. (Netherlands) were used in extruding both films and fibres. This is a long-chain semi-crystalline thermoplastic polymer with a density of 1.34 g/cm³, water absorption (equilibrium, 23 °C, 50% RH) of 0.30%, viscosity number of 85.0 cm³/g, melt temperature of 255 °C and processing temperature range between 265 - 295 °C.

Prior to extrusion, the PET pellets were dried in an oven for two days at 70 °C, after which pure *B. amyloliquefaciens* spores were homogeneously mixed with the pellets to obtain a 0.5% w/w concentration of spores to PET polymer. Finally this mixture was extruded in a Haake poly-drive extruder (Thermo Electronic Corporation, USA). The three different heating zones of the barrel were set at 265 °C for the feed stock (transport the material), 270 °C for plasticising (compression) and 275 °C for pumping (metering). The die was heated to 275 °C, while the screw speed and take-up velocity were set at 20 r.p.m. and 80 m/min respectively, resulting in a pressure of 0.5 MPa and film thickness of 0.8 mm. The average residence time during the extrusion was approximately one minute. The distribution and abundance of spores in the films were determined by Scanning Electron Microscopy (Joel Quanta 200 F FE-SEM). Samples for the SEM were prepared by placing them on a stub, where they were gold coated using a sputter coater afterwards (Balzers Union SKD 030).

#### Extrusion of spores in PET fibres

After drying the PET pellets as explained earlier, pure *B. amyloliquefaciens* spores were added by gravimetric dosing during spinning to obtain a 0 (control), 2, 4, 6, 8 and 10% w/w concentration of spores to PET polymer. This mixture was extruded into multi-filament fibres by a single screw extruder (General Extrusion Technology, China). The three different heating zones of the barrel were set at 280 ± 10 °C, 290 ± 5 °C and 295 ± 2 °C. The die was heated at 295 °C and the pressure was around 6.0 ± 0.2 MPa, while the average residence time was approximately 5 ± 0.5 minutes.

#### Resistance of extruded spores in films and fibres

The resistance of the extruded spores in the PET films and fibres was determined by growth/germination biological assay

### Table 1. Test parameters under which the *Bacillus amyloliquefaciens* spores were tested.

<table>
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</tr>
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</table>

### Table 2. Results for two-sample Kolmogorov-Smirnov tests of the effect of temperature, pressure and residence time on number of viable spores (n - number of replica); 1 - Control represents a control temperature of 21 °C, 2 - Control represents a control pressure of 0.1 MPa, 3 - Control represents a control time of 0 minutes.

<table>
<thead>
<tr>
<th>Test parameters</th>
<th>Amount of samples (n)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Temperature, °C</td>
<td>0.913</td>
<td></td>
</tr>
<tr>
<td>Pressure, MPa</td>
<td>0.286</td>
<td></td>
</tr>
<tr>
<td>Time, min.</td>
<td>0.517</td>
<td></td>
</tr>
</tbody>
</table>

### Table 3. Surviving spores in fibres after being sterilised by soaking in sodium hypochlorite (NaClO) solution, Dettol solution (5%) and in ethanol solution (96%) for 10 minutes each or by pasteurizing at 80 °C for 10 minutes (n - number of replicas).

<table>
<thead>
<tr>
<th>Concentration of spores, %</th>
<th>Amount of samples (n)</th>
<th>Sterilised (Mean Log CFU/g dry fibres)</th>
<th>Pasteurised (Mean Log CFU/g dry fibres)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>2</td>
<td>14 ± 0</td>
<td>14 ± 1</td>
<td>20 ± 1</td>
</tr>
<tr>
<td>4</td>
<td>20 ± 1</td>
<td>19 ± 1</td>
<td></td>
</tr>
</tbody>
</table>
which means that *B. amyloliquefaciens* spores that survived the extrusion processing conditions will form a colony after inoculation and be detected as viable. Before inoculation, the samples had to be sterilised to remove contamination due to the extrusion. Several sterilisation techniques like pasteurisation and sterilisation with chemicals can be used [18]. In order to test the most effective sterilisation method to use, two techniques were tested: pasteurisation and sterilisation with chemicals.

The samples were cut into small pieces to end up with 48 samples, each weighing about 85 mg. Half of them were then sterilised by soaking in Sodium hypochlorite (12% NaClO), in Dettol solution (5%) and in ethanol (96%) for 10 minutes each. The other half of the samples was sterilised by pasteurising in a water bath (Grant JB aqua 12) at 80 °C for 10 minutes. After each sterilisation process, the samples were dissolved in 1.0 ml of sterile physiological saline solution, then diluted and inoculated on nutrient agar plates and incubated at 40 °C for 24 hours. Finally the plates were visually checked for colony forming spots.

To generate a statistical equation that explains the relationship between the survival rate and concentration of extruded *B. amyloliquefaciens* spores, experimental results from the ‘sterilised’ column were used (Table 3).

## Results and discussion

### Effect of temperature

The effect of temperature on the heat resistance of *B. amyloliquefaciens* spores is given in Figure 1.a, showing a decrease in the number of viable spores with increasing temperature. The decrease in surviving spores means spores were killed during the heat treatment, increasing with a rise in temperature. Extreme dry heat can kill spores by damaging and mutating the DNA [19 - 21]. DNA damage is mostly due to loss of base through depurination [10, 22 - 24].

### Effect of pressure

The effect of pressure on the viability of *B. amyloliquefaciens* spores is shown in Figure 1.b. For the pressure range tested, from 0.1 to 1.0 MPa, pressure seemed to have no effect on the number of surviving spores. An explanation for the resistance of spores to pressure could be provided by DNA-binding proteins protecting the spore’s DNA, the repair of DNA damage during spore germination, and by the impermeability of the spore’s coat to high pressure [8, 9]. Additionally *B. amyloliquefaciens* spores were reported to resist pressures up to 1,700 MPa [26 - 28], which is much higher than the highest pressure tested (1.0 MPa). The pressure range tested was too low to kill the spores and therefore is not an important parameter in the extrusion of spores/PET films and fibres.

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![Figure 1](image1.png)

**Figure 1.** The effect of temperature (a) and pressure (b) on the viability of *Bacillus amyloliquefaciens* spores (four replicas each).

![Figure 2](image2.png)

**Figure 2.** The effect of residence time on the viability of *Bacillus amyloliquefaciens* spores; a) under pressure (1.0 MPa) and b) under high temperature (300 °C) (four replicas each).
B. amyloliquefaciens spores are well known to be resistant to high pressures [27 - 29], a pressure of 1.0 MPa is too low to cause any decrease in the survival of spores, even after 10 minutes of exposure. In contrast, the effect of the residence time at high temperatures is a linear decrease in the amount of surviving spores with increasing temperature (Figure 1a). This is in accordance with previous results, in that fewer spores of B. amyloliquefaciens survive with increasing temperature [11, 12], which means that any change made in the residence time or temperature will have a great effect on the number of spores that survive the process. The longer the spores are exposed to high temperatures, the more likely it becomes that the spores will not survive the processing.

**Statistical analysis**

The statistical significance of the temperature, pressure and residence time on the number of viable spores was determined using the two-sample Kolmogorov-Smirnov test, the results of which are given in in Table 2. This test compared the distribution of values for each parameter against the control values. The analysis showed that none of the parameters tested saw a significant difference in the viable spores (Table 2, p > 0.05), meaning that neither temperature, pressure, nor time had a significant influence on the number of spores that survived the process. This lack of significance may be attributed to the low operating pressure levels and short residence time, which were discussed earlier. Additionally high temperature treatment has been shown to increase the relative hydrophobicity of the spore surfaces due to the denaturing of spore coat proteins, resulting in an agglomeration of the spores, which increases their survival chances against extreme temperatures [20, 30 - 33].

**Extrusion of spores in PET films**

PET films of 0.8 mm thickness incorporated with B. amyloliquefaciens spores were successfully extruded in a stable process. The surface of the PET films was very characteristic, in that many spots were visible due to the inclusion of the spores in the film, as can be seen in Figure 3. This is a first indication of the feasibility to successfully incorporate bacteria spores in extruded polymer films.

The viability of the spores in the spore/PET films was tested by inoculation in nutrient medium and incubating for 24 h at 40 °C. This tests the survival of spores through germination/growth under favorable conditions [18]. The viability test results of the PET films are shown in Figure 4. The control sample, being pure PET film with no spores (Figure 4A), did not show any growth, whereas the sample extruded with 0.5% spores showed bacterial growth all around the polymer film (Figure 4B). The growth observed in Figure 4B confirms that B. amyloliquefaciens spores survived the extrusion parameters very well.

As discussed earlier, the resistance of B. amyloliquefaciens spores to PET film extrusion can be explained by three factors, namely the low operating pressures, the short residence time and the formation of spor agglomerates during extrusion. Several factors can cause the spores to agglomerate in the extruder: the hydrophobic nature of the spores, heat treatment and shear stress [34, 35]. An example of spore aggregation in the PET film is given in the SEM picture shown in Figure 5.

**Extrusion of spores in PET fibres**

The concentration of spores successfully incorporated in PET fibres was from 0 to 10% w/w. Concentrations higher than 10% w/w blocked the spinnerets and therefore extrusion could not take place.

The two sterilising techniques studied proved to be equally efficient for fibres before inoculation (Table 3). PET fibres extruded with a 2% spore concentration showed the smallest survival of spores,
while concentrations of 4, 6, 8 and 10% of spores showed almost the same survival rate. These results further confirm the resistance of *B. amyloliquefaciens* spores to melt extrusion process parameters.

### Quadratic equation

The equation developed to relate the survival rate of spores to the concentration of extruded spores was

\[
y = 1.714 + 5.82x - 0.42x^2
\]

where, \(y\) is the number of viable spores, \(x\) the concentration of compounded PET pellets, and \(x^2\) is the squared term for the concentration of compounded PET pellets. The fitted curve is represented in Figure 6 \((R^2 = 0.91)\).

This quadratic equation can be used to relate the survival rate of *B. amyloliquefaciens* spores with concentration of spores extruded in PET fibres at 265 - 300 °C temperature, 6.0 ± 0.2 MPa pressure and 5 ± 0.5 minutes residence time.

### Conclusion

In this study, we have demonstrated a suitable technique for testing the resistance of spores to extrusion process parameters (temperature, pressure and residence time). Results showed that none of the parameters tested had a significant effect on the survival of *Bacillus amyloliquefaciens* spores. Therefore, the technique demonstrated proved to be cheap, easy and fast, hence it can be used as a model system to study the biological response of spores to high temperature, pressure and residence time. It was also shown that spores can be successfully incorporated directly into PET polymer matrix during extrusion to develop textile bio-aggregates.

### Acknowledgement

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### References


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